

Discrete embryonic character variation uncovers hidden ecological adaptations in lacertid lizards

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Abstract

Embryogenesis is the first step in the ontogenetic life journey of any individual, and is thus a starting point for natural selection to cause evolutionary change. There are slight variations in the timing of embryonic development, known as heterochrony, which may eventually lead to major differences in adult anatomy. To test this hypothesis, the embryonic development of three closely related lizard species, *Darevskia armeniaca*, *Lacerta agilis*, and *L. viridis*, which are adapted to different habitats, was compared by analyzing discrete timing characters. Both intra- and interspecific variation was detected. The latter may be interpreted as embryonic pre-adaptations to later adult lifestyles, demonstrating that developmental penetrance manifests within a few million years. Traits with large intraspecific temporal variation, such as limb-related features, were susceptible to natural selection. In particular, the mountain-dwelling, climbing species *D. armeniaca* showed embryonic preadaptations by an early developing limb anlagen. This observation demonstrated interspecific variation, which was elusive in a previous comparative study based on purely metric data of developing limb lengths, and highlighted the importance of multiple data sources to draw robust conclusions about evolutionary change. Timing differences indicated unexplored ecological adaptations of the poorly understood lifestyle of these lizards. Thus, embryonic research provides a platform to explore superficially hidden evolutionary adaptations of all organisms on Earth.

KEYWORDS

developmental penetrance, embryogenesis, natural selection, organ development, reptiles, standard event system (SES)

1 | INTRODUCTION

Natural selection occurs not only in adults, but also throughout the ontogeny of every organism (Maier, 1999). For example, hatchlings of the European green lizard have an extraordinarily low chance of survival in their first year; their mortality rate increases up to 100% in years with bad conditions, while adults may live up to approximately ten years (Hill & Klepsch, 2008). One major source for selection is anatomical variation (Raff, 1996), resulting in major taxon diversification, as well as in fast radiation in lower taxonomic levels, such as ‘families’ or ‘genera’. As individual anatomic features form

during embryonic development, it is important to examine these for a comprehensive understanding of how evolutionary processes are linked to individual ontogeny. Werneburg et al. (2015) have argued that, in general, “the earlier an element appears during development, the more prominent it becomes in the adult because it has more time to grow and differentiate.” Thus, the later a structure emerges, the smaller or less complex it will appear. Such adaptive heterochrony implies, among other things, deep developmental penetrance, indicating that evolutionary adaptations are genetically integrated into embryonic development (Hamrick, 2001; Richardson, 1999). In this way, morphological features that are

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characteristic for adaptation in adults develop early and may become fully established should they be required directly after birth or hatching (Bickelmann et al., 2012).

A good model for these research questions are the true lizards, Lacertidae (Figure 1), which currently number over 300 species that “successfully radiated into a wide array of habitats and climate regions from subarctic tundra over temperate heath lands and forests, alpine meadows and Mediterranean maquis, steppe and gravel semi-deserts, and monsoonal rainforest to sandy dune systems in the desert” (Baeckens et al., 2015). This occurred in a relatively short time, geologically speaking, of approximately 85 million years (Kumar et al., 2017). To mirror this diversity, the intra- and interspecific differences in the ontogeny of three lacertid species adapted to different habitats was investigated: the Armenian rock lizard *Darevskia armeniaca*, sand lizard *Lacerta agilis*, and European green lizard *L. viridis*.

In a recent attempt to understand interspecific limb variation through the ontogeny of the three lacertid species, no significant difference among embryos, but only post-embryonic development of selected limb proportions was identified (Cordero et al., 2021). However, only selected metric traits were used in this approach, which did not account for internal anatomical specifications or differences in the general embryonic anatomy among the three species. The different nature of the data used in the present study may be better suited to distinguish changes in embryonic development.

Because development is a continuous process and traditional “normal embryonic staging” is not well suited for documenting variation (Richardson, 2022), the Standard Event System (SES) approach introduced by Werneburg (2009) was used, which uses discrete characters (Figure 2) to describe and compare intra- and interspecific differences in embryogenesis.

FIGURE 1 Embryos of different developmental periods and adults of the three species analyzed in this study. Photo of adult *Darevskia armeniaca*: Köhler, Babette (2019-08-11), via <https://www.inaturalist.org/photos/47933393>, CC BY-NC 4.0, accessed: 2022-04-11. Photo of adult *Lacerta agilis*: Böhringer, Friedrich (2007-08-30), via https://commons.wikimedia.org/wiki/File:M_Zauneidechse1_Edit1.jpg, CC BY-SA 3.0, accessed: 2022-04-11. Photo of adult *Lacerta viridis*: Uoaei1 (2014-04-12), via <https://commons.wikimedia.org/w/index.php?curid=39840199>, CC BY-SA 4.0, accessed: 2022-04-11



In this study, the following hypotheses will be investigated: Since no feeding or hunting specializations were mentioned in the literature for the three species, it was assumed that no heterochrony in embryonic jaw development would be identified. In contrast, because vision is a very important sense for lacertids, not only for foraging or avoiding predators, but also for intraspecific communication, Martin et al. (2014) have suggested “that the visual system and visual signals might co-evolve.” Thus, variations in eye genesis between species with different lifestyles were expected. Similarly, pronounced interspecific embryonic differences in limb development were expected among non-climbing or only occasionally climbing species. Some heterochrony in scale development, possibly related to microornamentation based on habitat preference was also anticipated, as suggested by Arnold (2002).

2 | MATERIAL AND METHODS

2.1 | Species

2.1.1 | The European green lizard *Lacerta viridis*

Numerous populations of the European green lizard *Lacerta viridis* are distributed throughout Europe, up to elevations of 2000 m above sea level. These lizards characteristically inhabit a shrubby habitat, such as that found on roadsides, forest edges, or clearings (Engelmann et al., 1986). Shrubs are of great importance to the species and are used both for sunbathing and protection from predators (Böhme, 1984). Although it is an occasional climber, *L. viridis* disappears when the habitat becomes overgrown, and the potential for thermoregulation in sun-exposed areas is lost (Kirmse, 1990). *L. viridis* does not occur in steppes or coniferous forests, and feeds on all types of insects, especially beetles, but also eats eggs, juvenile mammals, and small reptiles (Böhme, 1984).

With a snout-vent length of up to 13 cm and a very long tail that extends up to 40 cm, *L. viridis* is one of the largest lizards among

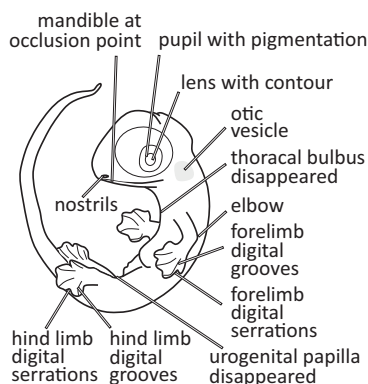


FIGURE 2 Selected standard event system (SES) characters coded in this study illustrated for a mid-term embryo (image modified from Yaryhin & Werneburg, 2017: fig. 1a). For more details on SES-characters see: https://en.wikipedia.org/wiki/Standard_Event_System [accessed: 2022-04-11]

European species (Figure 1, Engelmann et al., 1986). Adults of both sexes are colored shades of green, ranging from yellowish to blue, and sometimes have a dark pattern; males often have blue throats, particularly during the breeding season (Engelmann et al., 1986). Juveniles are brown, usually without a pattern, and blend in with their environment (Böhme, 1984). Böhme (1984) attributes the extremely variable incubation time for *L. viridis* (other authors mention a period between 50 and 100 days) to their large distribution area and the consequent variation in incubation parameters (Engelmann et al., 1986; Hill & Klepsch, 2008).

2.1.2 | The sand lizard *Lacerta agilis*

Distributed and common in almost all European countries, the sand lizard *Lacerta agilis* is one of the few reptiles that are described as synanthropic, meaning that it thrives in anthropogenic landscapes with structures, such as road and railway embankments, parks, cemeteries, and gardens (Böhme, 1984).

Many subspecies exhibit a wide variety of colors in all shades of green and brown and patterns from nearly solid to dots and stripes, with strong sexual dimorphism (Böhme 1984; Engelmann et al., 1986). *L. agilis* is a medium-sized lizard (Figure 1), reaching a snout-vent length of approximately 11 cm with a total length of up to 27.5 centimeters (Böhme, 1984; Engelmann et al., 1986).

The sand lizard feeds on arthropods and lizard eggs and occasionally exhibits cannibalistic behavior (Böhme, 1984). Similar to *L. viridis*, the incubation time of *L. agilis* is variable; known values range from approximately 32–36 days at very high temperatures (28°C) to over 60–75 days (Peter, 1904) or even 70–90 days (Märtens, 1999) at lower temperatures.

2.1.3 | The Caucasian rock lizard *Darevskia armeniaca*

The Caucasian rock lizard, *Darevskia armeniaca*, has been described and taxonomically reclassified several times. Consequently, there is some ambiguity in the nomenclature and common descriptions. Girnyk et al. (2018) state that the Caucasian rock lizard “originated through interspecific hybridization between the closely related sexual *Darevskia mixta* and *Darevskia valentini*,” which, according to Darevsky (1966), “apparently took place in forest refuges during the Quaternary glaciation of the Caucasus.” Today, the distribution of *D. armeniaca* extends from northwestern Turkey through South Georgia and northern Armenia to northwestern Azerbaijan (Bischoff, 2003). It was intentionally introduced into Ukraine in 1963 with the aim to “study the process of acclimatization and hybrid complexes of lizards” (Nekrasova & Kostiushev, 2016).

Bischoff (2003) has described its occurrence as linked to rocky habitats, such as rivers or roadbanks, at elevations from above 1350 m to above 2200 m. In contrast to other lacertids, purely female populations appear extraordinarily nonviolent towards each other (Karmyshev & Yaryhin, 2013). Adults are green or olive in color with a black pattern and a white or yellow belly (Figure 1), and hatchlings are

coppery with a green tail (Bischoff, 2003). Karmyshev and Yaryhin (2013) have reported an incubation period of 66 days.

2.1.4 | Outgroup comparison

For the outgroup comparison in the phylogenetic analysis, the black and white tegu *Salvator merianae* (Teiidae) was chosen because it is phylogenetically close to Lacertidae (Streicher & Wiens, 2017), and it has a more or less generalized lizard anatomy without limb reduction (e.g., snakes, amphisbaenians, and skinks) or large eyes (e.g., geckos), and very detailed embryonic data are available for this species (lungman et al., 2008).

2.2 | Embryonic sampling

To obtain embryonic material of the lacertid species, gravid females of all species were caught in their natural habitats in Ukraine from 2007 to 2011 (Figure 1). The capture of animals is allowed if they are not listed in the Red Data Book of Ukraine (Akimov, 2009). *Lacerta viridis* was added to the list in 2009, but was captured before that, in 2007. The female lizards were kept in terraria where they successfully laid their eggs. Animal husbandry was approved by the Ethics Committee of the Schmalhausen Institute of Zoology NAS of Ukraine. The eggs were incubated on moistened vermiculite at temperatures ranging from 21 to 23°C (Karmyshev & Yaryhin, 2013; Yaryhin & Werneburg, 2018). The maximum incubation times were 85 days, 65 days, and 66 days for *L. viridis*, *L. agilis*, and *Darevskia armeniaca*, respectively. For the latter, the authors mention “a rather large number of embryonic malformations (up to 7%), and twins were also found twice in the eggs”, which they account to their parthenogenic mode of reproduction (Karmyshev & Yaryhin, 2013). Because almost no genetic mixing by mating occurs in parthenogenetic species (Darevsky et al., 1978), acquired mutations accumulate in the lineage, causing an unusually high rate of malformed embryos. No malformed embryos of any species were analyzed in the present study, but data from embryos with delayed development were used if no other embryos were available. As gravid females were collected from the field, whether males were involved in reproduction cannot be stated with certainty. Nevertheless, it is important to note that neither more nor less intraspecific variation was detected in *D. armeniaca* when compared to both *Lacerta* species.

The embryos obtained were then fixed in formalin and stored in 4% buffered formalin, 100% methanol, or 70% ethanol. Some embryos were stained with phosphotungstic acid (PTA) to prepare them for another study using μ CT scans.

2.3 | Data sampling

2.3.1 | Photography

The specimens were photographed using a Leica Z16 Apo trinocular microscope (Leica Biosystems Technology, Stuttgart, Germany) equipped

with a camera and refined using Image-Pro Plus software (Media Cybernetics, Rockville, USA). If possible, dorsal, ventral, and lateral views were obtained, as proposed by Werneburg (2009) (some specimens were broken, twisted, or too small). An example is shown in Figure 1.

2.3.2 | Discrete character coding

To enable quantitative and qualitative comparison of inter- and intra-specific variation in embryogenesis, Werneburg (2009) has developed the “Standard System to Study Vertebrate Embryos” (SES). The system provides a comprehensive catalogue of more than 100 homologous external developmental characters in various complexes, such as the eye, pharyngeal arch, and limb morphology. By carefully examining every specimen and assigning observable characters, one receives a discrete, comparable description of an embryonic series, which may easily be supplemented by new characters or specimens, as shown in Figure 2. The SES has been expanded throughout the last few years by Werneburg et al. (2016) and the updated database can be accessed herein: https://en.wikipedia.org/wiki/Standard_Event_System (access: 2022-04-11). As initial development of embryos takes place in the oviduct, because of the sampling technique described above, there was only access to embryos of advanced development, so not all SES characters could be observed. Some characters described in the SES did not apply to reptiles or lacertids. Therefore, the following SES-characters were not documented: complexes A, B, D, U, V, W, X, Y, Z, G4, H6, I1, L23, L24, L29, L30, M11, N1-N4, S1-2, and T3 (Werneburg, 2009; Werneburg et al., 2016).

In the present study, 15 *Lacerta viridis* embryos, 33 *L. agilis* embryos, 36 *Darevskia armeniaca* embryos, and 28 *Salvator merianae* embryos taken from the literature (lungman et al., 2008) were examined. For complete information and SES data of the specimens, see Tables S1–S3.

2.3.3 | Data comparison among species and phylogenetic analysis

Despite the different incubation lengths given in the literature, the age of each specimen has been scaled by the maximum incubation time given by Karmyshev and Yaryhin (2013), which were obtained through controlled and constant conditions, to make comparisons of the timing of developmental characters possible. Werneburg et al. (2016) have discussed whether the birth and hatching of different animals may be homologous, which would be crucial for SES coding, and answered positively, as the breach of the amnion is a homologous event for all amniotes. In that case, it was decided against using birth as the maximal value, as mammals are very differently developed at birth; that is, on a spectrum of precocial to altricial. As lizards are precocial animals and basically ‘miniature adults’ apart from coloring, it was used here because the four species are similarly sized at hatching. This way, all events were scaled resulting in values between ‘0’ (i.e., egg lay) and ‘1’ (i.e., hatch) for every

developmental character in every species. Using conception as a starting point for comparison would have been a better proxy. However, all three compared species were taken from the field and stable life conditions of the females there could not be guaranteed. With fluctuating temperatures, development may last longer or be shorter (diapauses). As such, for the most reliable comparison in lizards, it was decided to consider egg laying as a starting point of compared development because the incubation conditions were controlled and stable.

To examine intraspecific variation, the earliest and latest values and the arithmetic mean for the timing of each character were compared. For the comparison of interspecific variation, the earliest value was used instead of the arithmetic mean because many characters persist for a long period of time after their first appearance, which affects the mean calculation. This procedure also circumvented the problem arising from the variable time the egg remained in the uterus until it was laid. Since each specimen only shows one continuous trait at a time within a complex, and trends within a complex were compared instead of individual traits, a robust developmental trend for each species was obtained. Individual specimens that were developmentally delayed owing to malformations were immediately conspicuous by external features and compared to conspecifics. Hence, for a proper interspecific comparison, only the earliest appearance of a character was chosen (see Werneburg et al., 2016).

With these values, a phylogenetic analysis using Mesquite version 3.61 (Maddison & Maddison, 2018), applying the continuous character and square change parsimony approach (Felsenstein, 1985) was performed (as in Germain and Laurin, 2009). This resulted in ancestral state reconstructions: the values for the ancestral timing of the characters for the 'genus' *Lacerta* and for the ancestor of all 'Lacertidae' (Table S4, Figures 3–6).

The time of evolutionary divergence of the four species was calculated using timetree.org (last access: 2021-10-01) from Kumar et al. (2017), which uses the best estimates from a variety of molecular studies. The calculation resulted in the following divergence times: *Lacerta agilis*/*L. viridis* = 21.7 million years, *Darevskia armeniaca*/*Lacerta* = 44.8 million years, *Salvator merianae*/Lacertidae = 168 million years.

For comparison, the ancestral sequences of Lacertidae and the individual sequences of the three lacertid species were plotted in four diagrams (Figures 3–6). The sequence data were sorted by the timing of character appearance in the lacertid ancestor.

3 | RESULTS

3.1 | Intraspecific variation in developmental sequence

Each of the species studied here exhibited some degree of intraspecific variation; that is, the same embryonic characters were observed to appear and/or disappear at different times during ontogeny.

Statistical tests on this variation were not performed because there were too few data points for a sufficient analysis. Therefore, this study represents an exploratory, rather than a quantitative assessment (as in [Parsi-Pour & Werneburg, 2019]), often the only approach to deal with rare embryological data. To estimate intraspecific variation, the earliest and latest occurrence and arithmetic mean of the timing of SES traits were presented for the three lacertids (Figures 3–5, Table S4). Developmental sequences were sorted according to the reconstructed ancestral states of all three species observed in this study.

The Armenian rock lizard *Darevskia armeniaca* (Figure 3, Table S1) was highly variable in the temporal sequence of the visual and auditory organ systems, particularly in the extremities. As discussed in more detail in the Methods section, many specimens of *D. armeniaca* had malformations or were generally delayed in their development; for example, they seemed 'normally developed', but were older than expected. One noteworthy example is DA80, the only specimen in which digital grooves were observed in the forelimbs ('L15') and hind limbs ('L16'). Therefore, the values for these characters were higher than those for 'L19. Finger' or 'L20. Toe' and were treated as outliers in intra- and inter-specific comparisons. When compared to the development of the reconstructed ancestral state, *D. armeniaca* often seemed delayed at the beginning of development, but completed its growth as the earliest of the species.

A few specimens were available for the green lizard *Lacerta viridis* (Figure 4, Table S2). In several cases, SES characters could only be observed in one embryo, and many were absent altogether, making the dataset for this species the sparsest. This also affected observable intraspecific variation. Nevertheless, the development of sensory organs exhibited a great deal of temporal variation, similar to limb growth. The oldest specimen (LV8) had no scales, but this was probably because of its poor preservation. With a few exceptions, the development of *L. viridis* was delayed compared to that of the ancestral state reconstruction.

In contrast, the sand lizard *L. agilis* (Figure 5, Table S3) began embryonic development earlier than the reconstructed ancestral state, but growth appeared to slow as it progressed, and organogenesis was completed later. For *L. agilis*, many specimens were well distributed throughout embryogenesis, so there were few problematic characters. 'P4, the mandibular arch bud midline eye' appeared to be delayed, but as only one specimen displayed this character it was treated as an outlier. The development of the hind extremities occurred later than that of the fore extremities.

3.2 | Interspecific variation and evolutionary change

The earliest values for all three species, as well as ancestral state reconstruction, are shown in Figure 6, illustrating the heterochronic embryonic development.



FIGURE 3 Intraspecific variation in the timing of embryological development of the Armenian rock lizard *Darevskia armeniaca*, scaled by hatch = 1, and the reconstructed ancestral lacertid condition (Lacertidae). In addition to the four illustrated body region categories (face, sensory organs, limbs, and scales), also early ontogenetic characters are listed for which no ancestral lacertid states could be calculated because the outgroup is missing this information. Abbreviations of characters such as “P1. Mandibular arch bud” or “L04. Hind limb bud” refer to the official SES numeration and SES character abbreviation system, as introduced by Werneburg (2009) and Werneburg et al. (2016) and as summarized at: https://en.wikipedia.org/wiki/Standard_Event_System [accessed: 2022-04-11]

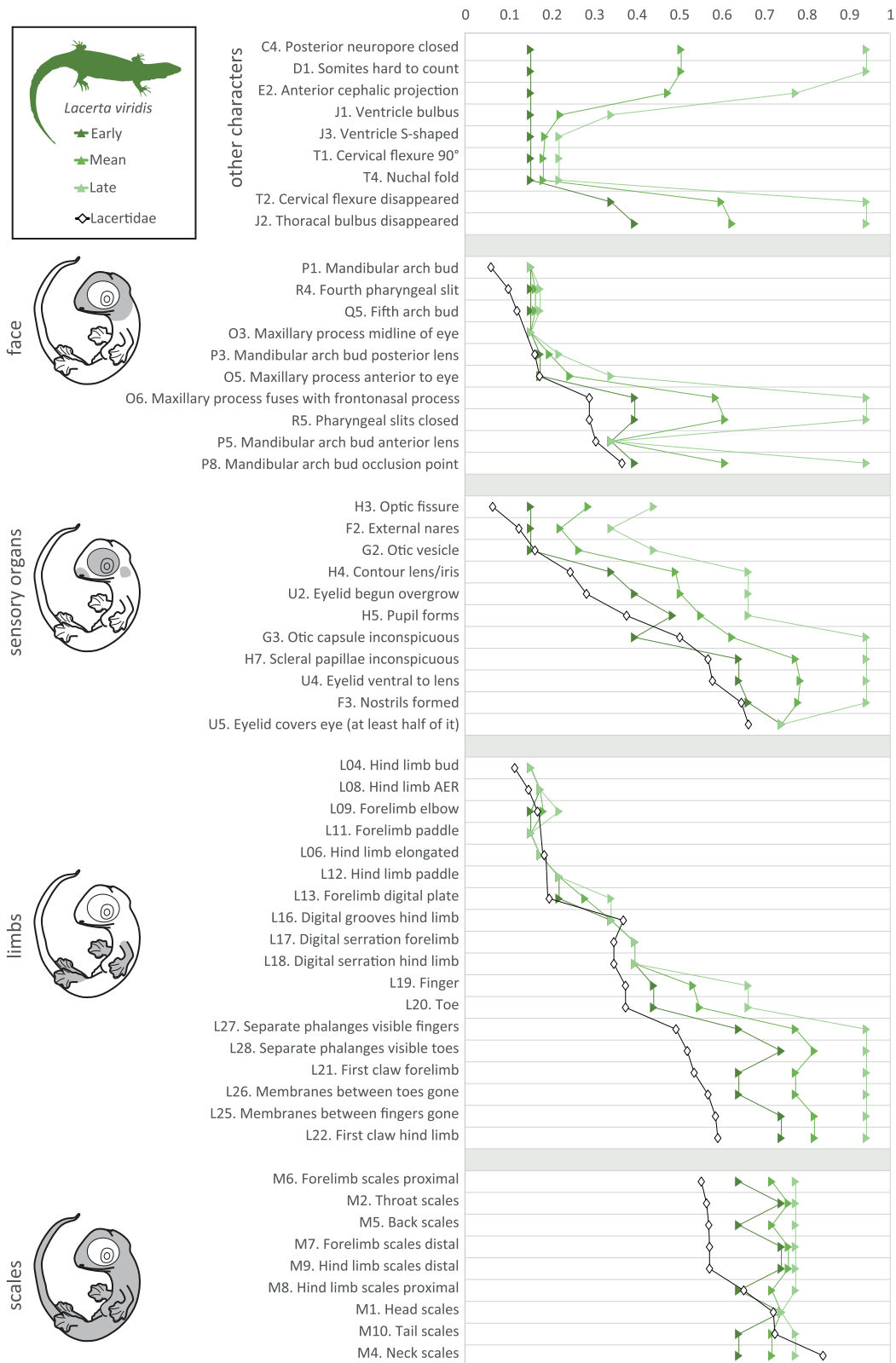


FIGURE 4 Intraspecific variation in the timing of embryological development of the green lizard *Lacerta viridis*, scaled by hatch = 1, and the reconstructed ancestral lacertid condition (Lacertidae). For abbreviations, see the legend of Figure 3

The ontogeny of the face, that is, the snout and pharyngeal region, is dominated by the growth of the jaw, which originates from the first pharyngeal (mandibular) arch that forms “a dorsal maxillary

and a ventral mandibular process’ (Werneburg, 2009). These processes move from the posterior to the anterior of the eye, where the former fuses with the nasal region and both extend anteriorly until



FIGURE 5 Intraspecific variation in the timing of embryological development of the sand lizard *Lacerta agilis*, scaled by hatch = 1, and the reconstructed ancestral lacertid condition (Lacertidae). For abbreviations, see the legend of FIGURE 3

the occlusion point of the maxilla and mandible is reached (Werneburg 2009). Initially, a similar pattern was recognizable among the studied species; at the time of oviposition, some pharyngeal slits

and arches were developed and rapidly closed and dislimned as the maxilla and mandible moved anteriorly. Contrary to our first hypothesis of simultaneous ‘jaw development’ in all three lacertid species,

closure of pharyngeal slits, fusion of the maxillary process with the nasal region, and finally jaw closure was first achieved by *Darevskia armeniaca*, in the first third of embryogenesis. In contrast, both *Lacerta*

species completed jaw formation at approximately the same time in the second third of the incubation period, with *L. viridis* occurring a little earlier.

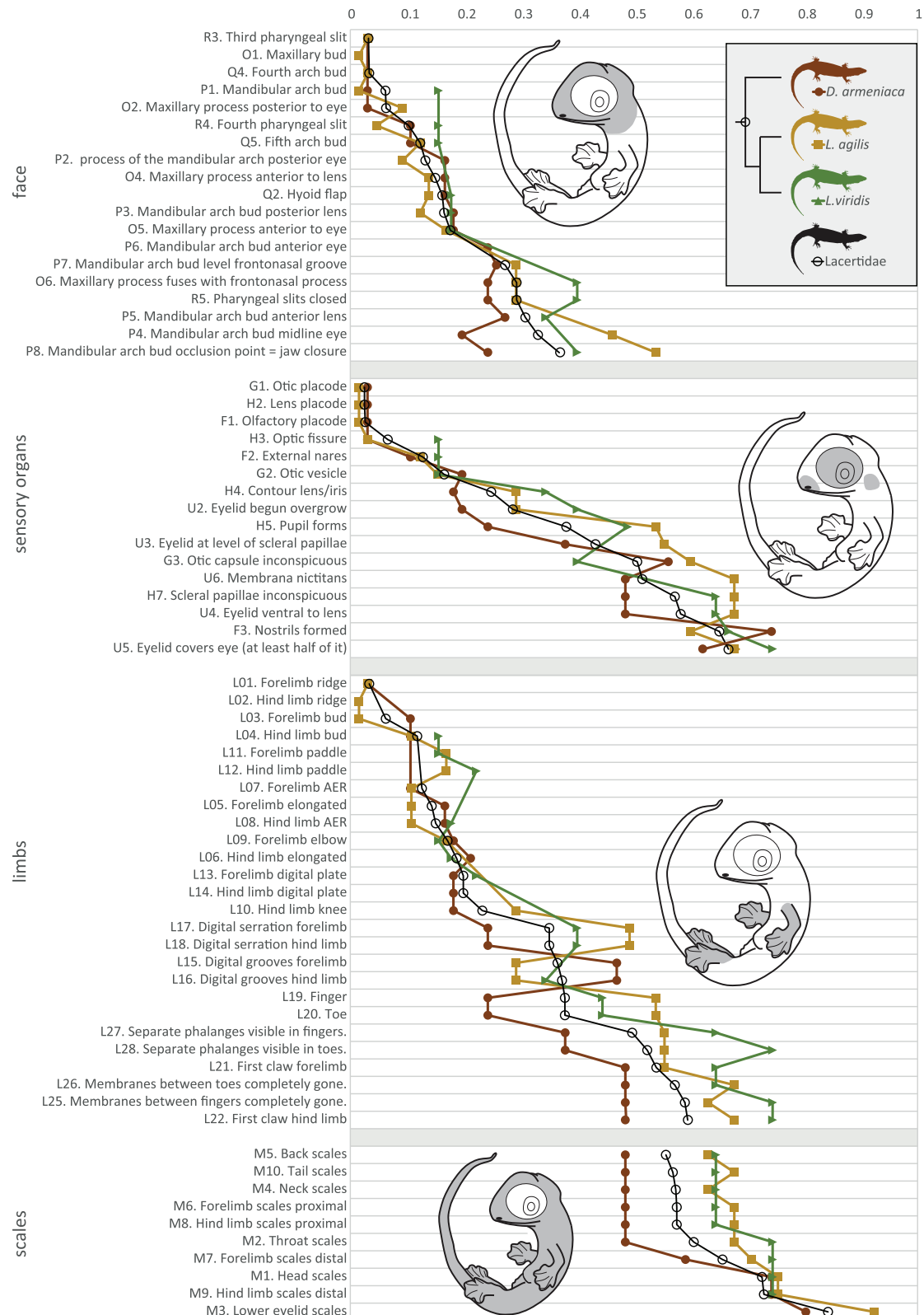


FIGURE 6 Interspecific variation in the earliest appearance of embryological characters (face, sensory organs, limbs, and scales) in the three species, *Darevskia armeniaca*, *Lacerta viridis*, and *L. agillis*, scaled by hatch = 1, and the reconstructed ancestral lacertid condition (Lacertidae). For abbreviations, see the legend of FIGURE 3

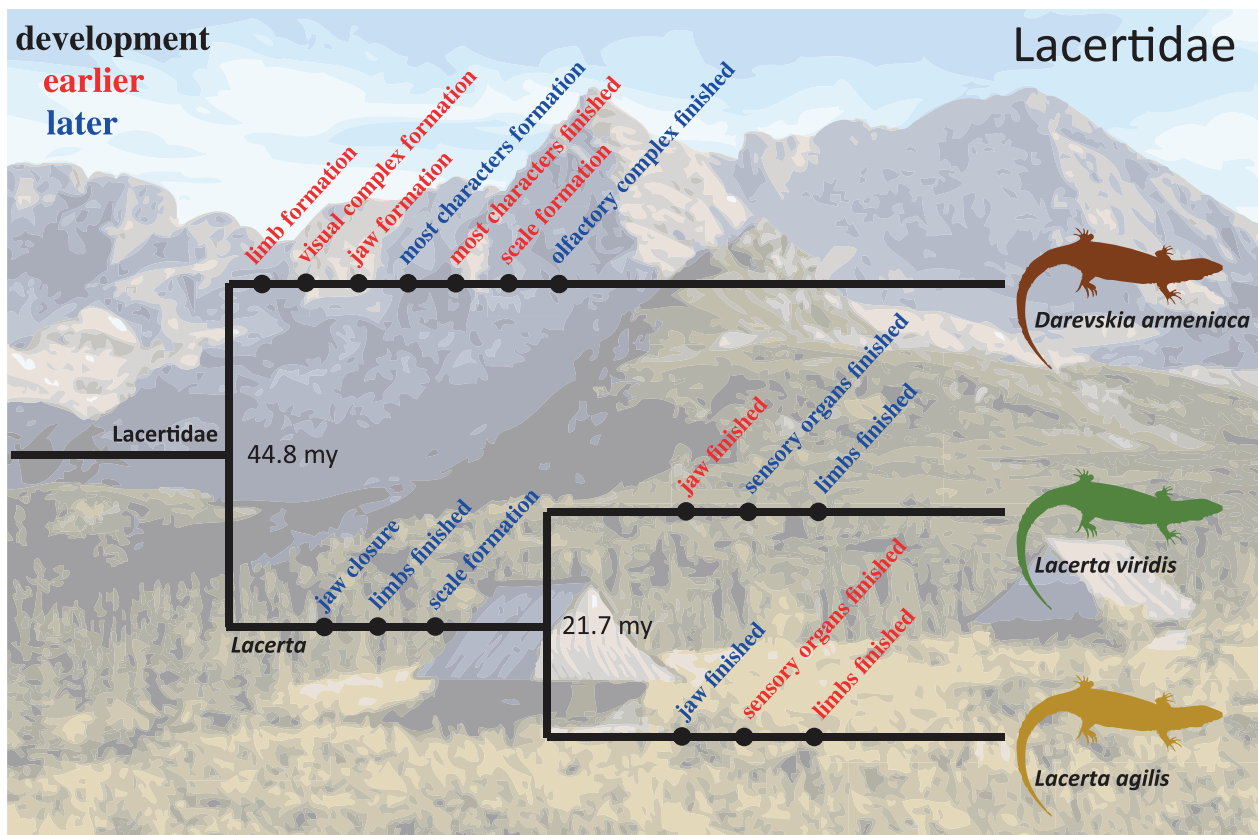


FIGURE 7 Habitat range and phylogenetic relationship, including the time of evolutionary divergence for the species *Darevskia armeniaca*, *Lacerta viridis*, and *L. agilis*. Time in million years. Derived characters on the tree represent a summary of the findings of this study. Red writing indicates earlier, blue writing indicates later character appearance in ontogeny. Characters on the tree present summaries of one or more standard event system (SES)-character development(s) that are illustrated in Figures 3 and 4. Landscape illustration in the background is a personal adobe illustrator artwork based on a photo indicating the habitat for the respective species with *Darevskia armeniaca* living in mountainous environments. *Lacerta viridis* inhabits a characteristically shrubby habitat, such as roadsides, forest edges, or clearings. *L. agilis* thrives in anthropogenic landscapes with structures, such as road and railway embankments, parks, cemeteries, and gardens

Regarding the development of facial senses, interspecific differences were expected in the visual and nasal complex. Accordingly, the auditory complex developed quite similarly in all three species. In contrast, but consistent with our second hypothesis, the visual system exhibited interspecific differences in its development, with *D. armeniaca* displaying both the direct visually related characters and those of the later-emerging eyelid complex much earlier in development. *D. armeniaca* was the last and *L. agilis* was the first species to have fully formed nostrils.

In accordance with the expected interspecific variation in limb development, the most striking heterochrony was observed in the character complexes associated with locomotion. Limb development began immediately after oviposition in all species and continued into the second third of embryogenesis. While *D. armeniaca* completed the development halfway through the incubation period, *L. agilis* took just until the last third and *L. viridis* unexpectedly took the longest.

When examining scale development, the observed interspecific differences were consistent with our fourth hypothesis, as it was expected to find heterochrony in this character complex. Scale onset seemed to be connected to the completion of limb development, and consequently,

D. armeniaca started and finished scale growth first. This followed the general pattern of all three species, according to which scales grew only after the body had formed and grown. Scales first appeared on the back and neck, whereas the scales on the lower eyelid grew last.

4 | DISCUSSION

Intraspecific variation among lacertid species was observed, particularly in limb, facial, and scale development. These variations were mirrored in interspecific differences, as they were most likely related to postnatal adaptations, implying developmental penetrance.

It should be noted that the ‘early equals importance’ rule (Huxley, 1932; Mehnert, 1897; Mehnert, 1898) does not necessarily mean that the early appearance of a character correlates with the size of a fully developed anatomical feature. It may also be related to structural differentiation; for example, increased complexity of internal anatomy, such as diversification of muscles or ossification (Werneburg & Sánchez-Villagra, 2009). As discussed by Werneburg et al. (2015), this also allows for the execution of ontogenetic

functions that occur only for a limited period of time (see also Werneburg et al., 2013).

Cordero et al. (2021) have concluded that *Lacertini* differs in limb development only after hatching by comparing the quantitative and metrical data of very similarly sized animals. In the present study, discrete and qualitative data of morphology were compared. Therefore, the results of the two studies do not necessarily contradict each other, but are also not strictly comparable. Furthermore, not only late limb development, where whole limb length is measured (Cordero et al., 2021), but also the earliest states of limb differentiation have been examined, including limb ridge, limb bud, and digital serration, as well as specific differentiations, such as finger formation.

4.1 | Facial development

4.1.1 | Formation of the jaw

All three lacertid species studied herein feed on a similar, mainly insectivorous diet, and no specialization has been mentioned in the literature for any of the species (Böhme, 1984, 1984). Therefore, no embryological differentiation of jaw formation was expected. Surprisingly, the data showed that *Darevskia armeniaca* completed closure of the maxillary and mandibular processes, as well as pharyngeal slits, considerably earlier than the other species studied during the first third of embryogenesis (Figure 7). Further research on adult and hatchling dietary specifications might reveal a link to the observed interspecific differences. However, since the formation of facial features is one of the processes that begin earliest in organogenesis (Werneburg 2009), and most specimens in this study were further developed, these results must be verified by comparing younger specimens.

4.1.2 | Development of facial sensory organs

Regarding the development of the external sensory organs of the head, noticeable intra- and interspecific variation was observed in eye formation (Figure 7). Research on visual sense in lacertids has only just begun, but as Martin et al. (2014) have suggested, it may be of much greater importance than previously thought, including for intraspecific communication. For example, some species display visual signals in the ultraviolet or infrared range, which requires retinal specialization (Martin et al., 2014). In this study, *D. armeniaca* achieved eye development earlier than both *Lacerta* species, strongly suggesting developmental penetrance of a specialized visual system.

Interestingly, *D. armeniaca* was the last, and *L. agilis* was the first species to exhibit fully formed nostrils. As suggested by Baeckens et al. (2015), chemical signaling in lizards may be subject to strong sexual selection and may be particularly important to males. Considering that the Armenian rock lizard is a parthenogenic species, the delayed formation of the nostril suggests that olfaction might play a minor role in this species.

4.2 | Locomotion

The most striking difference among the three lacertid species is the way they move in their different habitats (Figure 7). Therefore, limb development was expected to vary embryonically, and, indeed, both intraspecific and interspecific variation was observed. *Darevskia armeniaca*, as a mountain dweller and consequently an actively climbing species, completed limb development earlier than *Lacerta viridis*, *L. agilis*, and also earlier than the reconstructed ancestral sequence of lacertid development. Notably, *L. viridis*, as an occasional climber in the lower shrub layer, completed limb development later than *L. agilis*, which remains on the ground in the herbaceous layer and does not climb (Böhme, 1984). Apart from length (Cordero et al., 2021), there are no obvious external anatomical differences in the limbs of adults between all lizard species. However, based on its climbing behavior, it is very likely that *D. armeniaca* has different and longer internal anatomical specializations in the limbs, including myology and degree and mode of ossification (Bischoff, 2003).

4.3 | Scales

To protect the animal's body, scales must be fully developed before hatching. However, because scales do not grow or multiply, shedding is required to make room for the growing body (Spearman, 1973). Because the embryo cannot shed its skin (Spearman, 1973), it is crucial for it to complete growth processes earlier than scale formation. Interestingly, the onset of scale growth was observed in areas that do not experience changes later in embryogenesis, such as the back and neck. These conditions limit the applicability of the 'early equals importance rule'.

In the present study, scale formation started at the same time as the completion of limb development in all species. However, while *Lacerta agilis* and *L. viridis* appeared to develop scales similarly in the last third of embryogenesis, *Darevskia armeniaca* finished scale formation earlier (Figure 7).

According to Arnold (2002), scale microornamentation in lacertids is linked to the habitat and behavior of the species and exhibits two opposite characteristics. The more ancestral mode would enhance smoothness to avoid soiling and facilitate movement in narrow crevices, which also increases light reflection (Arnold, 2002). A more derived scale surface would reduce smoothness to avoid attracting predators and does appear dorsally in *L. viridis* and *L. agilis* (Arnold, 2002). Reduced smoothness is associated with "dry habitats or when they live away from the ground, both situations where soiling is unlikely to be a great problem" (Arnold, 2002). It is not known which sort of microornamentation appears in *D. armeniaca*, but given their close association with rocks and their montane habitat at high elevations, both the ancestral smooth and the derived reflection-reducing pattern seem likely. Comparing the embryological data, it seemed likely that as both *Lacerta* species completed scale growth later and have more derived scale microornamentation, *D. armeniaca*

probably has a more ancestral one. This would also fit their lifestyle, hiding in rock crevices.

4.4 | Developmental penetrance

We hypothesized there would be intra- and interspecific differences in embryonic development, the latter representing an embryonic pre-adaptation to the later lifestyle and anatomical adaptation of adult lizards of *Darevskia armeniaca*, *Lacerta agilis*, and *L. viridis*. Using the SES approach developed by Werneburg (2009) to make continuous embryonic differentiation comparable, interspecific variation was identified, particularly in limb development, but also in scale, sensory, and facial development. Therefore, it seems that, although separated for only 44 million years, changes in adult anatomy have clearly penetrated into embryogenesis.

The main results concern the differences in limb development. In this regard, *D. armeniaca* developed limbs considerably earlier, suggesting a specialized adaptation to its rocky habitat and climbing lifestyle. Since no external anatomical differences of the limbs were evident in adults, this should be investigated further by studying their extremities, especially internal morphological differentiation.

In all species, the development of scales occurred within a very short interval from their first appearance and their completion just before hatching, although they are very important to both the hatchlings and adults (Spearman, 1973). This is due to the fact that the whole body must be developed before scales appear, as after scales cover the animal, growth is only possible by shedding, which is a well-developed feature in squamates (Spearman, 1973). Nevertheless, there were differences among the species, with *D. armeniaca* developing scales first, which was most likely related to adaptations, such as scale microornamentation.

Surprisingly, a large difference was found in the timing of facial development, namely, in the growth of the jaw and in the genesis of the eye and its eyelids. In both cases, *D. armeniaca* exhibited the most rapid development, suggesting a developmental penetrance of specific adaptations that have yet to be discovered. It is very likely that there is some sort of specification in *D. armeniaca* for both the diet and visual system, perhaps related to its relatively small distribution range and rocky habitat.

The early development of *D. armeniaca* indicated mature hatchlings compared to that of other lacertid lizards. This may be particularly advantageous in extreme conditions given their montane habitat, where only a short period between long winters is suitable for lizard activity, but young hatch only in autumn (Nekrasova & Kostiusshyn, 2016).

4.5 | Conclusions

Reptiles are a useful model to address fundamental questions regarding evolutionary mechanisms because most lay eggs, which allows for good access to numerous embryos at different developmental stages

for research. In addition, some of their clades are extremely diverse and radiate into “empty niches” within a few decades (Stuart et al., 2014). One example is Lacertidae, in which rapid diversification supported by developmental penetrance has manifested itself in as little as 44 million years (Kumar et al., 2017).

Intraspecific variation was observed in the timing of ontogeny prior to hatching during the development of the face, sensory organs, locomotor system, and scales. This suggested sufficient variation to cause natural selection at the earliest stages of ontogeny, thereby enabling speciation by following the ‘early equals importance’ rule.

The observation of concordant interspecific variation further supports our hypotheses (Figure 7). In *D. armeniaca*, an earlier jaw development, an earlier formation of the visual complex, a later completion of the olfactory complex, a very early limb, and scale development was observed when compared to those of the ancestral lacertid condition. In general, most characters showed a late onset of formation, but an early completion. Compared to that of the ancestral Lacertidae condition, *Lacerta* was characterized by later jaw closure, formation of sensory organs, and limb and scale development. Compared to that of the ancestral *Lacerta* condition, *L. viridis* showed later formation and earlier completion of jaw and limb development. Its sensory organ and scale development were delayed compared to those of *Lacerta*. Compared to that of the ancestral *Lacerta* condition, *L. agilis* showed earlier formation and later completion of jaw development, while sensory organ development took place earlier. In contrast, scale development was delayed and the limbs developed similar to the ancestral *Lacerta* condition (Figure 7).

These results are inconsistent with previous studies on lacertid limb development, in other words, they were elusive in previous comparative studies based on purely metric data (Cordero et al., 2021). This highlights the importance of multiple data sources to draw robust conclusions about evolutionary change. Therefore, the stable observation of intra- and interspecific patterns of variation across whole-body development was interpreted as an indication of the importance of using discrete data sources and a more qualitative approach to draw conclusions about evolutionary change. Especially since development is a continuous process that cannot be measured using quantitative measurements alone. This conclusion was based on these observations that although no dietary or hunting specialization has been mentioned in the literature, heterochrony was identified in the embryonic jaw development of the three species. In addition, the differences in the development of the facial sensory system, as well as the limbs and scales, suggested deep developmental penetrance of adaptations to slightly different lifestyles and habitats, which might account for the success of the diverse lacertids.

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CONFLICT OF INTEREST

None.

AUTHOR CONTRIBUTIONS

Conceptualization: IW, OY, XS.

Collected specimens: OY.

Analyses: XS, OY.

Wrote paper: XS, IW.

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SUPPORTING INFORMATION

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specimen	DA40	DA80	DA36	DA5	DA11	DA9	DA12	DA13	DA15	DA16	DA17	DA76
egg lay date	21.06.15	22.06.11	22.06.15	22.06.11	28.06.11	28.06.11	28.06.11	28.06.11	28.06.11	28.06.11	28.06.11	28.06.11
date of fixation	16.07.15	22.07.11	23.07.15	27.07.11	03.08.11	05.08.11	05.08.11	07.08.11	15.08.11	17.08.11	19.08.11	26.08.11
days incubated	26	31	32	35	37	39	39	41	49	51	53	60
fixation				F	F	F	F	F	F	F	F	F
place	UDZ		UDZ	UDZ	UDZ	UDZ	UDZ	UDZ	UDZ	UDZ	UDZ	UDZ
C4. Posterior neuropore closed	0.393939394	0.46969697	0.484848485	0.53030303	0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
D1. Somites hard to count	0.393939394	0.46969697	0.484848485	0.53030303	0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
E2. Anterior cephalic projection	0.393939394	0.46969697		0.53030303								
E3. Head projection disappeared								0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
F1. Olfactory placode												
F2. External nares		0.46969697		0.53030303								
F3. Nostrils formed									0.742424242	0.772727273	0.803030303	0.909090909
G1. Otic placode												
G2. Otic vesicle	0.393939394	0.46969697		0.53030303								
G3. Otic capsule inconspicuous					0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
H2. Lens placode												
H3. Optic fissure		0.46969697		0.53030303								
H4. Contour lensiris	0.393939394	0.46969697										
H5. Pupil forms	0.393939394											
H7. Scleral papillae inconspicuous			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
J1. Ventricle bulbus		0.46969697		0.53030303								
J2. Thoracic bulbus disappeared	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
J3. Ventricle S-shaped				0.53030303								
L1. Forelimb ridge												
L3. Forelimb bud												
L5. Forelimb elongated												
L7. Forelimb AER												
L9. Forelimb elbow	0.393939394	0.46969697	0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L11. Forelimb paddle												
L13. Forelimb digital plate												
L15. Digital grooves forelimb		0.46969697										
L17. Digital serration forelimb												
L19. Finger:												
L21. First claw forelimb			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M. Membranes between the fingers are completely gone.			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L27. Separate phalanges are visible in the fingers:	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L2. Hind limb ridge												
L4. Hind limb bud												
L6. Hind limb elongated												
L8. Hindlimb AER												
L10. Hind limb knee	0.393939394	0.46969697	0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L12. Hind limb paddle												
L14. Hind limb digital plate												
L16. Digital grooves hind limb		0.46969697										
L18. Digital serration hind limb												
L20. Toe												
L22. First claw hind limb			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L26. Membranes between the toes are completely gone.			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
L28. Separate phalanges are visible in the toes	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M1. Head scales									0.742424242	0.772727273	0.803030303	0.909090909
M2. Throat scales			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M3. Lower eyelid scales											0.803030303	0.909090909
M4. Neck scales			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M5. Back scales			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M6. Forelimb scales proximal			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M7. Forelimb scales distal					0.590909091	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M8. Hind limb scales proximal			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
M9. Hind limb scales distal									0.742424242	0.772727273	0.803030303	0.909090909
M10. Tail scales			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
O1. Maxillary bud												
O2. Maxillary process posterior to eye												
O3. Maxillary process midline of eye												
O4. Maxillary process anterior to lens				0.53030303								
O5. Maxillary process anterior to eye												
O6. Maxillary process fuses with frontonasal process	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
P1. Mandibular arch bud												
P2. process of the mandibular arch posterior eye												
P3. Mandibular arch bud posterior lens				0.53030303								
P4. Mandibular arch bud midline eye												
P5. Mandibular arch bud anterior lens												
P6. Mandibular arch bud anterior eye												
P7. Mandibular arch bud level frontonasal groove												
P8. Mandibular arch bud occlusion point = jaw closure	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
Q1. Second arch bud												
Q2. Hyoid flap				0.53030303								
Q3. Third arch bud												
Q4. Fourth arch bud				0.53030303								
Q5. Fifth arch bud												
R1. First pharyngeal slit												
R2. Second pharyngeal slit												
R3. Third pharyngeal slit												
R4. Fourth pharyngeal slit				0.53030303								
R5. Pharyngeal slits closed	0.393939394		0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
T1. Cervical flexure 90°	0.393939394			0.53030303								
T2. Cervical flexure disappeared			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
T4. Nuchal fold				0.53030303								
U1. Lower lid appears												
U2. Eyelid begun overgrow												
U3. Eyelid at level of scleral papillae	0.393939394											
U4. Eyelid ventral to lens			0.484848485		0.560606061	0.590909091	0.590909091	0.621212121	0.742424242	0.772727273	0.803030303	0.909090909
U5. Eyelid covers eye (at least half of it)								0.621212121	0.742424242		0.803030303	0.909090909

Table S2. SES data for all *Lacerta viridis* embryos, scaled by maximum incubation time (compare chapter 4.2)

specimen	LV16	LV13	LV15	LV8	LV10	LV9	LV7	LV12	LV11	LV5	LV1	LV3	LV4	LV2	LV6
egg lay date	25.05.07	25.05.07	25.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07	27.05.07
date of fixation	07.06.07	09.06.07	13.06.07	23.06.07	01.07.07	05.07.07	09.07.07	11.07.07	13.07.07	17.07.07	23.07.07	25.07.07	01.08.07	04.08.07	19.08.07
days incubated	14	16	20	31	36	40	44	46	48	52	58	60	67	70	85
fixation	F	F	F	F	F	F	F	F	F	F	F	F	F	F	F
place	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE	UDE
C4. Posterior neuropore closed	0.15555556	0.17777778	0.22222222	0.34444444	0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
D1. Somites hard to count	0.15555556	0.17777778	0.22222222	0.34444444	0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
E2. Anterior cephalic projection	0.15555556	0.17777778	0.22222222	0.34444444	0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
E3. Head projection disappeared															
F1. Olfactory placode															
F2. External nares	0.15555556	0.17777778	0.22222222	0.34444444											
F3. Nostrils formed												0.68888889	0.74444444	0.77777778	0.84444444
G1. Otic placode															
G2. Otic vesicle	0.15555556	0.17777778	0.22222222	0.34444444		0.44444444									
G3. Otic capsule inconspicuous					0.4		0.48888889	0.51111111	0.53333333				0.74444444	0.77777778	0.84444444
H2. Lens placode															
H3. Optic fissure	0.15555556	0.17777778	0.22222222	0.34444444	0.4	0.44444444									
H4. Contour lens/iris				0.34444444	0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778		0.68888889			
H5. Piggy forms				0.34444444	0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778		0.68888889			
H7. Scleral papilla inconspicuous												0.64444444	0.74444444	0.77777778	0.84444444
I1. Ventricles bulbous	0.15555556	0.17777778	0.22222222	0.34444444											
J2. Thoracic bulbus disappeared					0.4		0.48888889	0.51111111	0.53333333				0.74444444	0.77777778	0.84444444
J3. Ventricles S-shaped	0.15555556		0.22222222												
L1. Forelimb ridge															
L3. Forelimb bud															
L5. Forelimb elongated															
L7. Forelimb AER															
L8. Forelimb elbow	0.15555556	0.17777778	0.22222222												
L11. Forelimb paddle	0.15555556														
L13. Forelimb digital plate			0.22222222	0.34444444											
L15. Digital grooves forelimb															
L17. Digital serration forelimb					0.4										
L18. Finger					0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889		0.74444444	0.77777778	0.84444444
L21. First claw forelimb															
L27. Separate phalanges are visible in the fingers												0.64444444	0.74444444	0.77777778	0.84444444
L2. Hind limb ridge															
L4. Hind limb bud	0.15555556														
L6. Hind limb elongated		0.17777778													
L8. Hindlimb AER		0.17777778													
L10. Hind limb knee															
L12. Hind limb paddle			0.22222222												
L13. Hind limb digital plate															
L16. Digital grooves hind limb				0.34444444											
L18. Digital serration hind limb					0.4										
L20. Toe					0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889		0.74444444	0.77777778	0.84444444
L22. First claw hind limb															
L26. Membranes between the toes are completely gone												0.64444444	0.74444444	0.77777778	0.84444444
L28. Separate phalanges are visible in the toes												0.64444444	0.74444444	0.77777778	0.84444444
M1. Head scales															
M2. Throat scales															
M3. Lower eyelid scales															
M4. Neck scales												0.64444444	0.74444444	0.77777778	0.84444444
M5. Back scales												0.64444444	0.74444444	0.77777778	0.84444444
M6. Forelimb scales proximal												0.64444444	0.74444444	0.77777778	0.84444444
M7. Forelimb scales distal												0.64444444	0.74444444	0.77777778	0.84444444
M8. Hind limb scales proximal												0.64444444	0.74444444	0.77777778	0.84444444
M9. Hind limb scales distal												0.64444444	0.74444444	0.77777778	0.84444444
M10. Tail scales												0.64444444	0.74444444	0.77777778	0.84444444
O1. Maxillary bud															
O2. Maxillary process posterior to eye															
O3. Maxillary process midline of eye	0.15555556														
O4. Maxillary process anterior to lens															
O5. Maxillary process anterior to eye	0.17777778	0.22222222	0.34444444												
O6. Maxillary process fuses with frontonasal process					0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
P1. Mandibular arch bud	0.15555556														
P2. process of the mandibular arch posterior to eye															
P3. Mandibular arch bud posterior lens	0.17777778	0.22222222													
P4. Mandibular arch bud midline eye															
P5. Mandibular arch bud anterior lens				0.34444444											
P6. Mandibular arch bud anterior eye															
P7. Mandibular arch bud level frontonasal groove															
P8. Mandibular arch bud occlusion point = jaw closure					0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
Q1. Second arch bud															
Q2. Hyoid flap	0.17777778	0.22222222													
Q3. Third arch bud															
Q4. Fourth arch bud															
Q5. Fifth arch bud	0.15555556	0.17777778													
R1. First pharyngeal slit															
R2. Second pharyngeal slit															
R3. Third pharyngeal slit															
R4. Fourth pharyngeal slit	0.15555556	0.17777778													
R5. Pharyngeal slits closed					0.4	0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
T1. Cervical flexure 90°	0.15555556	0.17777778	0.22222222	0.34444444		0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
T2. Cervical flexure disappeared				0.34444444		0.44444444	0.48888889	0.51111111	0.53333333	0.57777778	0.64444444	0.68888889	0.74444444	0.77777778	0.84444444
T4. Nuchal fold	0.15555556	0.17777778	0.22222222												
U1. Lower lid appears															
U2. Eyelid begun overgrow					0.4	0.44444444	0.48888889	0.51111111	0.53333333			0.68888889			
U3. Eyelid at level of scleral papillae															
U4. Eyelid ventral to lens												0.64444444	0.74444444	0.77777778	0.84444444
U5. Eyelid covers eye (at least half of it)													0.74444444		

Table S4. Earliest, latest, and mean occurrence of SES data for all species, including outgroup and reconstructed, ancestral sequence

	<i>Darevskia armeniaca</i>			<i>Lacerta viridis</i>			<i>Lacerta agilis</i>			<i>Salvator merianae</i>	<i>Lacerta</i>	<i>Lacertidae</i>
	early	mean	late	early	mean	late	early	mean	late			
C3. Anterior neuropore closed										0.05		
C4. Posterior neuropore closed	0.03030303	0.37920875	0.90909091	0.15555556	0.50888889	0.94444444	0.01538462	0.96923077	0.45780886	0.05	0.46039176	0.41151531
D1. Somites hard to count	0.10606061	0.38917749	0.90909091	0.15555556	0.50888889	0.94444444	0.01538462	0.96923077	0.45780886	0.08333333	0.46240872	0.41782645
D6. 21-25 somite pairs										0.05		
E1. Head bulbus										0.06666667		
E2. Anterior cephalic projection	0.03030303	0.28058361	0.53030303	0.15555556	0.47777778	0.77777778	0.01538462	0.73846154	0.33969231	0.05	0.39435271	0.33244193
E3. Head projection disappeared	0.62121212	0.76969697	0.90909091	0.15555556	0.26888889	0.44444444	0.01538462	0.96923077	0.82692308	0.03	0.75867173	0.66387859
F1. Olfactory placode	0.03030303	0.03030303	0.03030303				0.01538462	0.09230769	0.05230769	0.06666667	0.04332274	0.04332274
F2. External nares	0.10606061	0.25974026	0.53030303	0.15555556	0.225	0.34444444	0.12307692	0.49230769	0.22615385	0.16666667	0.22665189	0.2351986
F3. Nostrils formed	0.74242424	0.80681818	0.90909091	0.66666667	0.78333333	0.94444444	0.6	0.96923077	0.75897436	0.25	0.764177	0.74932306
G1. Otic placode	0.03030303	0.10101010	0.16666667				0.01538462	0.13846154	0.08923077	0.05	0.19604048	0.09153131
G2. Otic vesicle	0.19669697	0.33164983	0.53030303	0.15555556	0.26888889	0.44444444	0.15384615	0.53846154	0.32527473	0.06666667	0.29640776	0.29497269
G3. Otic capsule inconspicuous	0.56060606	0.69886364	0.90909091	0.4	0.62857143	0.94444444	0.6	0.96923077	0.75897436	0.23333333	0.68422135	0.66387859
H2. Lens placode	0.03030303	0.10101010	0.16666667				0.01538462	0.12307692	0.07692308	0.05	0.18586697	0.08586697
H3. Optic fissure	0.03030303	0.24825175	0.53030303	0.15555556	0.29074074	0.44444444	0.03076923	0.53846154	0.22657343	0.06666667	0.25306797	0.24116856
H4. Contour lens/iris	0.18181818	0.3083779	0.46969697	0.34444444	0.49583333	0.66666667	0.29230769	0.53846154	0.44615385	0.1	0.44192045	0.38002281
H5. Pupil forms	0.24242424	0.32878788	0.39393939	0.48888889	0.55555556	0.66666667	0.53846154	0.73846154	0.63956044	0.16666667	0.55354592	0.45984277
H7. Scleral papillae inconspicuous	0.48484848	0.67508418	0.90909091	0.64444444	0.77777778	0.94444444	0.67692308	0.96923077	0.8	0.05	0.7320913	0.73982641
J1. Ventricle bulbus	0.03030303	0.25833333	0.53030303	0.15555556	0.225	0.34444444	0.01538462	0.49230769	0.15589744	0.05	0.19604048	0.20794553
J2. Thoracic bulbus disappeared	0.33333333	0.56493506	0.90909091	0.4	0.62857143	0.94444444	0.53846154	0.96923077	0.73393665	0.35	0.65899794	0.61161399
J3. Ventricle S-shaped	0.03030303	0.21380471	0.53030303	0.15555556	0.18888889	0.22222222	0.01538462	0.15384615	0.0951049	0.05	0.15189041	0.17295404
L01. Forelimb ridge	0.03030303	0.03030303	0.03030303				0.03076923	0.03076923	0.03076923	0.06666667	0.0341015	0.0341015
L02. Hind limb ridge	0.10606061	0.13636364	0.16666667				0.01538462	0.04615385	0.03076923	0.06666667	0.09947884	0.09947884
L03. Forelimb bud	0.10606061	0.15151515	0.16666667	0.15555556	0.15555556	0.15555556	0.10769231	0.16923077	0.13626374	0.08333333	0.14524158	0.14381925
L05. Forelimb elongated	0.16666667	0.16666667	0.16666667				0.10769231	0.16923077	0.14153846	0.1	0.14979895	0.14979895
L06. Hind limb elongated	0.21212121	0.21212121	0.21212121	0.17777778	0.17777778	0.17777778				0.1	0.18739669	0.18739669
L07. Forelimb AER	0.10606061	0.14646465	0.16666667				0.10769231	0.16923077	0.13846154	0.11666667	0.1404411	0.1404411
L08. Hindlimb AER	0.16666667	0.18933934	0.21212121	0.17777778	0.17777778	0.17777778	0.10769231	0.16923077	0.13626374	0.11666667	0.16027743	0.16271029
L09. Forelimb elbow	0.18181818	0.4315376	0.90909091	0.15555556	0.18518519	0.22222222	0.16923077	0.96923077	0.86153846	0.18333333	0.41909062	0.40990709
L10. Hind limb knee	0.18181818	0.42319749	0.90909091				0.29230769	0.96923077	0.68615385	0.18333333	0.52513707	0.52513707
L11. Forelimb paddle				0.15555556	0.15555556	0.15555556	0.16923077	0.16923077	0.16923077		0.16239316	0.16239316
L12. Hind limb paddle				0.22222222	0.22222222	0.22222222	0.16923077	0.16923077	0.16923077		0.1957265	0.1957265
L13. Forelimb digital plate	0.18181818	0.21969697	0.27272727							0.16666667	0.24746584	0.24746584
L14. Hind limb digital plate	0.18181818	0.21969697	0.27272727							0.16666667	0.24746584	0.24746584
L15. Digital grooves forelimb	0.46969697	0.46969697	0.46969697				0.29230769	0.46153846	0.37692308	0.18333333	0.40422097	0.40422097
L16. Digital grooves hind limb	0.46969697	0.46969697	0.46969697	0.34444444	0.34444444	0.34444444	0.29230769	0.46153846	0.37692308	0.23333333	0.37183946	0.39559031
L17. Digital serration forelimb	0.24242424	0.28353535	0.36363636	0.4	0.4	0.4	0.49230769	0.49230769	0.49230769	0.25	0.4211883	0.36803584
L18. Digital serration hind limb	0.24242424	0.28353535	0.36363636	0.4	0.4	0.4	0.49230769	0.49230769	0.49230769	0.25	0.4211883	0.36803584
L19. Finger	0.24242424	0.32575758	0.39393939	0.44444444	0.53703704	0.66666667	0.53846154	0.96923077	0.73393665	0.3	0.5883372	0.48795409
L20. Toe	0.24242424	0.32575758	0.39393939	0.44444444	0.53703704	0.66666667	0.53846154	0.96923077	0.73393665	0.3	0.5883372	0.48795409
L21. First claw forelimb	0.48484848	0.67508418	0.90909091	0.64444444	0.77777778	0.94444444	0.53846154	0.96923077	0.75589744	0.4	0.74694278	0.70458805
L22. First claw hind limb	0.48484848	0.67508418	0.90909091	0.64444444	0.77777778	0.94444444	0.67692308	0.96923077	0.79020979	0.35	0.77929113	0.72196271
L25. Membranes between the toes are completely gone	0.48484848	0.67508418	0.90909091	0.64444444	0.77777778	0.94444444	0.67692308	0.96923077	0.70332967	0.45	0.77306212	0.72363914
L27. Separate phalanges are visible in the fingers	0.37878788	0.62258953	0.90909091	0.64444444	0.77777778	0.94444444	0.53846154	0.96923077	0.75589744	0.4	0.74021174	0.6835244
L28. Separate phalanges are visible in the toes	0.37878788	0.62258953	0.90909091	0.64444444	0.77777778	0.94444444	0.53846154	0.96923077	0.75589744	0.4	0.75909206	0.6952897
M01. Head scales	0.48484848	0.67508418	0.90909091	0.74444444	0.74444444	0.74444444	0.74444444	0.96923077	0.87179487	0.45	0.8000161	0.78273337
M02. Thorax scales	0.48484848	0.67508418	0.90909091	0.74444444	0.74444444	0.74444444	0.67692308	0.96923077	0.78179489	0.5	0.75536771	0.71538961
M03. Lower eyelid scales	0.8030303	0.85060606	0.90909091				0.92307692	0.96923077	0.94871795	0.6	0.87833558	0.87833558
M04. Neck scales	0.48484848	0.67508418	0.90909091	0.64444444	0.72222222	0.77777778	0.63076923	0.96923077	0.77032967	0.6	0.73390571	0.70759069
M05. Back scales	0.48484848	0.67508418	0.90909091	0.64444444	0.72222222	0.77777778	0.63076923	0.96923077	0.77032967	0.35	0.7236519	0.69023262
M06. Forelimb scales proximal	0.48484848	0.67508418	0.90909091	0.64444444	0.72222222	0.77777778	0.67692308	0.96923077	0.78846154	0.45	0.73828395	0.70196706
M07. Forelimb scales distal	0.59090909	0.73989899	0.90909091	0.74444444	0.76111111	0.77777778	0.70769231	0.96923077	0.81076923	0.5	0.77369958	0.74763898
M08. Hind limb scales proximal	0.48484848	0.67508418	0.90909091	0.64444444	0.72222222	0.77777778	0.67692308	0.96923077	0.78846154	0.45	0.73828395	0.70196706
M09. Hind limb scales distal	0.74242424	0.80681818	0.90909091	0.74444444	0.76111111	0.77777778	0.75384615	0.96923077	0.87179487	0.6	0.80820433	0.79064265
M10. Tail scales	0.48484848	0.67508418	0.90909091	0.64444444	0.72222222	0.77777778	0.67692308	0.96923077	0.78106509	0.35	0.73292568	0.69307449
O1. Maxillary bud							0.01538462	0.07692308	0.04230769	0.06666667		
O2. Maxillary process posterior to eye	0.03030303	0.06818182	0.10606061				0.09230769	0.10769231	0.1	0.08333333	0.08403065	0.08403065
O3. Maxillary process midline of eye				0.15555556	0.15555556	0.15555556	0.12307692	0.12307692	0.12307692		0.19318553	0.30787499
O4. Maxillary process anterior to lens	0.16666667	0.28787879	0.53030303				0.13846154	0.16923077	0.15	0.11666667	0.15018184	0.15018184
O5. Maxillary process anterior to eye	0.18181818	0.24242424	0.31818182	0.17777778	0.24814815	0.34444444	0.16923077	0.46153846	0.3158462	0.16666667	0.27417095	0.25800002
O6. Maxillary process fuses with frontonasal process	0.24242424	0.47907648	0.90909091	0.4	0.59012346	0.94444444	0.29230769	0.96923077	0.69795711	0.18333333	0.61288573	0.54591108
P1. Mandibular arch bud	0.03030303	0.06818182	0.10606061	0.15555556	0.15555556	0.15555556	0.01538462	0.07692308	0.04230769	0.06666667	0.11402531	0.14616026
P2. process of the mandibular arch posterior eye	0.16666667	0.18181818	0.21212121				0.09230769	0.10769231	0.1	0.16666667	0.14295799	0.14295799
P3. Mandibular arch bud posterior lens	0.18181818	0.35606061	0.53030303	0.17777778	0.2	0.22222222	0.12307692	0.16923077	0.14505495	0.18333333	0.17395824	0.17700439
P4. Mandibular arch bud midline eye	0.196											